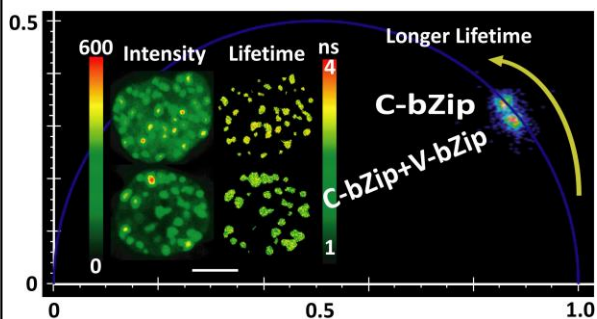
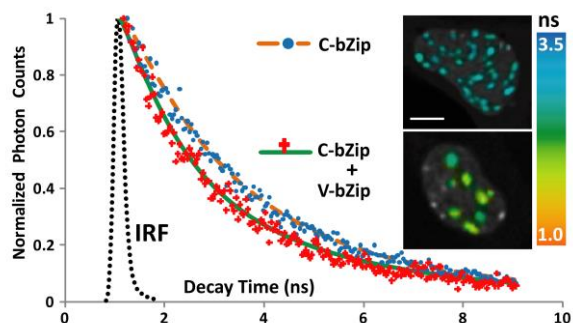


Multi-Photon tissue FRET in traumatic axonal injury. Energy transfer is consistent with BAD (Alexa488)-Bcl-xl (Alexa555) heterodimerization. Differentiation 71: 528-541, 2003



FLIM-FRET Microscopy to investigate proteins dimerizing in the nucleus.

Top: Time correlated single photon counting (TCSPC) method (Becker & Hickl).
Bottom: Frequency-domain technique (ISS).
Nature Protocols 6 (9), 1324-1340, 2011

Systems and live model cell line - transfected with above proteins will be available at workshop for the practical.

Faculty

Dr. A. Periasamy, University of Virginia
Workshop Coordinator, ap3t@virginia.edu

Dr. R. N. Day, Indiana University
Workshop Coordinator, rnday@iupui.edu

Dr. M. Barroso, Albany Medical College

Dr. K. Berland, Emory University

Dr. M. Börsch, Jena University

Dr. J. N. Demas, University of Virginia

Dr. A. Kenworthy, Vanderbilt University

Dr. F. Schaufele, University of California, SF

Dr. S. Vogel, NIAA, NIH

Dr. Jin Zhang, Johns Hopkins University

Guest Lecturers

Dr. M. Stanley, Chroma Tech.

Dr. P. So, MIT

For more info:

<http://www.kcci.virginia.edu/workshop/workshop2015>

FEES: \$2,200 non-profit organizations
\$2,600 for-profit organizations
(Includes lodging, breakfast, lunch, dinner, lecture materials)

Contact:

Horst Wallrabe – hw5m@virginia.edu

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Phone: 434-982-4869

15th Annual Workshop on FLIM and FRET Microscopy Imaging Protein- Protein Interactions March 7-11, 2016

Topics Includes

VFPs, Biosensors, FRAP, FCS, anisotropy, homo FRET, Inter- and Intramolecular Interactions, FRET data acquisition and analysis, participants presentation.



W.M. Keck Center for Cellular Imaging,
University of Virginia
www.KCCI.virginia.edu/workshop/workshop2016

AIM

The W.M. Keck Center for Cellular Imaging (KCCI), a university imaging center at the University of Virginia, is sponsoring an advanced practical course on Förster (fluorescence) resonance energy transfer (FRET) and fluorescence lifetime imaging (FLIM) microscopy.

Attendees are expected to be familiar with the basics of fluorescence microscopy. The curriculum, after a brief introduction to the principles of fluorescence, microscopy, fluorophores, FRET and FLIM, will concentrate on the practical aspects, hands-on individual instruction at the instruments followed by data analysis and interpretation.

Lectures and after dinner problem-solving discussions will address questions of fluorophore choices, the most suitable systems to achieve specific research objectives, qualitative vs. quantitative analysis and many more related subjects. Participants will also be introduced to a unique image processing and analysis software (PFRET).

10+ different and advanced microscopy systems will be available for a maximum of 25 students. With 13 internationally recognized faculty in attendance, there is ample opportunity to interact with experts formally or informally.

Live-cell specimens are provided.

Participant's own specimens are welcome.

PROGRAM SCHEDULE

Day 1 (12 Noon – 9 PM)*

- Introduction to workshop
- Basics of Fluorescence, FRET, FLIM, microscope choices
- Meet the experts from Leica, Olympus, Zeiss, Nikon, Chroma Tech, IdeaElan, Applied Precision, Lumen Dynamics, Becker & Hickl, Boston Electronics, ISS, Lambert.

Days 2-5 (8:30 AM – 12 PM)*

Short lectures and Q&A on the subjects:

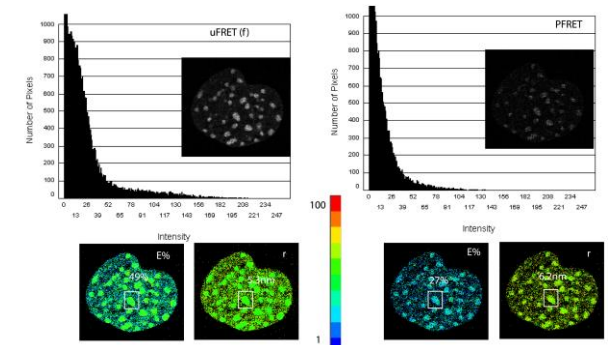
- Confocal/spectral wide-field FRET
- FLIM-FRET
- Fluorophore pairs for FRET/FLIM-FRET,
- Protein fusions
- Imaging live/fixed cells & tissue
- Spectroscopy FRET in suspensions
- Bacterial FRET

Days 2-5 (1 PM – 9 PM)*

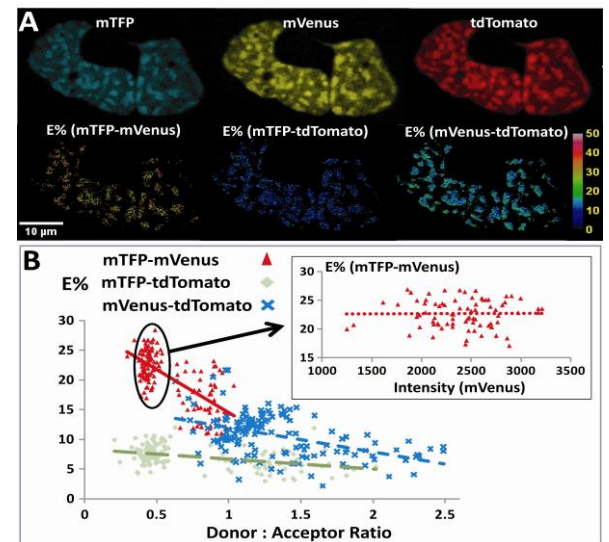
Hands-on practical instruction on various systems, data analysis, special demos and general problem-solving discussions on

- Anisotropy / Homo-FRET
- TIRF-FRET
- Spectrofluorometer FRET
- FLIM analysis: Fitting and Phasor plots
- Single-molecule FRET
- Fluorescence Correlation / Cross-correlation Spectroscopy
- FRAP
- Working on your instrument of choice after formal curriculum ends

***including breakfast, breaks, lunch and dinner.**



**Comparing uFRET (uncorrected for spectral bleed-through - SBT) with PFRET (Processed FRET, corrected for SBT).
This PFRET correction software will be available for workshop participants.**



**An expanded PFRET software analyzes 3 FRET-interacting, labeled proteins simultaneously in live cells – a Keck Center for Cellular Imaging development.
Biophys. J. 99, 1274-1283, 2010**

Participating Instrument Companies

Carl Zeiss, Leica Microsystems, Olympus, Nikon, ISS, Becker & Hickl, Boston Electronics, Chroma Tech, Lumen Dynamics, Semrock